

TRANSDERMAL DELIVERY OF FENOPROFEN: PREPARATION, EVALUATION AND *IN VITRO* RELEASE

Manjula D*¹, Shabaraya AR² and Somashekar Shyale³,

¹Dayananda Sagar College of Pharmacy, Bangalore, Karnataka, India.

²Srinivas College of Pharmacy, Mangalore, Karnataka, India.

³H.S.B.P.V.T. college of Pharmacy, Maharashtra, India.

Article Received on
29 Oct 2014,

Revised on 23 Nov 2014,
Accepted on 18 Dec 2014

*Correspondence for

Author

Manjula D

Dayananda Sagar College
of Pharmacy, Bangalore,
Karnataka, India.

ABSTRACT

The present study aimed at preparation and evaluation of fenopropfen transdermal drug delivery systems using various polymers like ERS100, ERL 100, HPMC and EC, in order to avoid its hepatic first pass metabolism and GI disturbance. The transdermal patches of fenopropfen were prepared by solvent casting method on surface of mercury, dibutyl phthalate (DBT) was used as plasticizer, d-limonene as permeation enhancer. The various formulations were evaluated for their physical appearance, thickness uniformity, weight uniformity, drug content and water vapour transmission (WVT). The *in vitro*

studies revealed that the addition of hydrophilic component (HPMC polymer) to an insoluble film former tends to enhance drug release. Further the transdermal therapeutic systems that showed better permeation were subjected for stability studies at 37⁰ C and 45⁰ C, which were found to be stable. Finally it was concluded that the above transdermal systems could be useful to treat chronic pain and inflammation in arthritis.

KEYWORDS: Fenopropfen, transdermal delivery, arthritis.

INTRODUCTION

Generally NSAID's are advised to reduce pain and inflammation in the treatment of arthritis which is one of the most affecting syndrome seen in geriatric patients. Usually conventional dosage forms like tablets and capsules are prescribed for the relief of inflammation and pain which are associated with arthritis.^[1] But these conventional dosage forms causes gastro intestinal disturbances and fluctuation in the drug level there by making the patient to suffer over dosage.^[2] In order to reduce the GI disturbance and to improve the bioavailability of the

drug, certain novel dosage forms are being investigated. Transdermal drug delivery systems (TDDS) are designed to support the passage of drug substances from the surface of skin, through its various layers, into the systemic circulation. Their advantages over conventional dosage forms include improved patient compliance, avoidance of gastric irritation and first-pass effect and controlled therapeutic responses.^[3]

Fenopfen, is one among the NSAID's used as anti inflammatory, analgesic and antipyretic. It is used in the treatment of osteoarthritis, rheumatoid arthritis and ankylosing spondylitis.^[4] Though it is rapidly being absorbed after oral administration, it undergoes significant first-pass metabolism. It has a very short half life of about 2-3 h and is associated with gastrointestinal side effects like nausea, gastric irritation etc. To extend drug action, to improve delivery of drug into systemic circulation, the present study was undertaken with the aim to develop and evaluate transdermal films of fenopfen using various polymers like ERL 100, ERS 100, HPMC and EC, plasticizer like DBP.^[5] Further the *in vitro* drug release was studied using d-limonene and oleic acid as permeation enhancers which belong to diterpene and fatty acids respectively. The formulation that showed promising result was further subjected for *in vivo* studies.

MATERIALS AND METHODS

Fenopfen was procured from D. K. Pharma, Mumbai, ERL 100, ERS 100, HPMC E-15 (Supra labs), EC (Himedia labs), d-limonene (Rolex chemical industries), Oleic acid (Qualigens fine chemicals) and all other chemicals obtained were of analytical grade. *In vitro* and *in vivo* experiments were carried out under the strict compliance of institutional animal ethical committee protocol bearing registration number 557/02/C/CPCSEA.

Drug Polymer Interaction by FT-IR^[6]

To investigate any possible interaction between the drug and the utilized polymer, IR spectrum of pure drug and its physical mixture with ERS 100, ERL 100, HPMC were carried out by using FT-IR (Perkin elmer 1600 series USA) the range selected was from 400cm^{-1} to 4000cm^{-1} .

Preparation of Transdermal Patches

Matrix type transdermal patches containing fenopfen were prepared by solvent evaporation technique (Mundane et al⁷ method) using different ratios of Eudragit RL 100(ERL 100) or Eudragit RS 100(ERS 100) and HPMC E 15. The formulae is depicted in table 1. The

polymers were weighed in requisite ratios by keeping the total polymer weight 2.0 gms and allowed for swelling for about 6 hrs in solvent mixture (1:1 ratio of dichloromethane, methanol). Propylene glycol was incorporated as plasticizer (30 % w/w of polymer) and d-limonene as penetration enhancer. Then specific quantity of drug was dissolved in alcohol and then added to the polymeric solution. The solutions were stirred for 20 minutes using magnetic stirrer and poured within a glass bangle of 5 cm diameter on mercury surface in petridish. The rate of evaporation of the solvent was controlled by inverting cut funnel over petridish. After 24 hrs the dried patches were taken out and stored in a dessicator. All the patches contained 5% d-limonene as penetration enhancer. The ratio of drug to polymer was 1:5 and it was kept constant for all the formulations. The formula is depicted as in table 1.

Table 1: Formulation table of various monolithic transdermal patches containing fenoprofen.

S.No.	Ingredients (mg/ml)	Formulation Codes									
		F1	F2	F3	F4	F5	F6	F7	F8	F9	F10
1.	Fenoprofen	50	50	50	50	50	50	50	50	50	50
2.	ERL 100	250	200	150	100	50	-	-	-	-	-
3.	ERS 100		-	-	-	-	250	200	150	100	50
4.	HPMC	-	50	100	150	200	-	50	100	150	200
5.	Propylene glycol	0.058	0.058	0.058	0.058	0.058	0.058	0.058	0.058	0.058	0.058
6.	d-limonene	0.003	0.003	0.003	0.003	0.003	0.003	0.003	0.003	0.003	0.003
8.	Dichloro methane	3	3	3	3	3	3	3	3	3	3
9.	Methanol	3	3	3	3	3	3	3	3	3	3

Evaluation of Formulated Patches

Physical Appearance

All the transdermal patches were visually inspected for colour, clarity, flexibility and smoothness.

Weight Uniformity

The dried patches were weighed on afcoset digital balance. The average of five observations were taken.

Thickness Uniformity^[8]

Film thickness was measured by a screw gauge at five different random points on the patch. The average of five observations were taken.

Water Vapour Transmission Studies^[9] (WVT)

Glass vials of equal diameter were used as transmission cells. These cells were washed and dried in an oven. About 1 gm of fused calcium chloride was taken in the cells and patch of area equivalent to brim of vial (1.36 cm^2) was fixed with the help of an adhesive. The cells were weighed accurately and kept in a close dessicator containing saturated solution of potassium bromide (200 ml). the humidity inside the dessicator was measured by a hygrometer and it was found to be 84% RH. The cells were taken out and weighed after 1, 2, 3, 4, 5, 6 and 7 days of storage. The WVT was calculated by taking the difference in the weight of the patches before and at regular intervals of 24 hrs for a total period of seven days.

Drug Content Uniformity^[10]

Transdermal films of 1 sq.cm area was cut into small pieces and transferred into 100 ml volumetric flask. 25 ml of methanol was added and shaken for 4 hours to extract the drug. Finally, suitable dilutions were made using phosphate buffer pH 7.4 and absorbance was measured at 272 nm.

In Vitro* Permeation Across the Rat Abdominal Skin*Preparation of Rat Abdominal Skin^[11]**

The swiss albino rats were sacrificed by decapitation. The fresh abdominal skin was excised from swiss albino rats weighing 170-190 gm. The abdominal skin of excised hairless rat skin was separated along the epidermal junction and was kept under a steam of 60°C water for exactly 50 seconds. The heat- treated skin was cleared of its subcutaneous fatty substances and immediately kept in normal saline solution to flatten and smooth. This step maintained the integrity and viability of the skin.

Permeation Studies

Vertically assembled Keshary-Chien^[12] diffusion cells having down stream volume of 50 ml was used. The above skin was mounted on the diffusion cell and receiver compartment was filled with 50 ml phosphate buffer of pH 7.4 and the temperature was maintained at 37°C . The samples were withdrawn every hour (replaced with 1 ml fresh buffer to maintain sink condition) and their concentrations were measured in UV-spectrophotometer at 272 nm.

1. Stability Studies^[13]

The stability studies were conducted at 37°C and 45°C for both (F5 and F10) films to investigate the influence of physical parameters and drug content of the films. Both the films

were exposed to selected temperatures of 37⁰ C and 45⁰ C in two different hot air ovens. Transdermal patch of area 19.63 cm² were wrapped individually in butter paper and placed in petridish in the oven for a period of four weeks. The sample of 1 cm² area of the film was cut from each formulation and it was analyzed for the drug content at the end of every week. Average of triplicate readings were taken.

RESULTS AND DISCUSSION

The IR spectra of pure drug and its physical mixtures with polymers (ERS 100, ERL 100 and HPMC) has showed replication of characteristic peaks of N-H stretch, C-H aromatic stretch, NO₂ stretch, C-O-C stretch, C-H aliphatic stretch and OH stretch respectively as in IR of pure drug. The results revealed no interaction between drug and polymers hence indicated their compatibility. Fig 1, 2, 3 and 4.

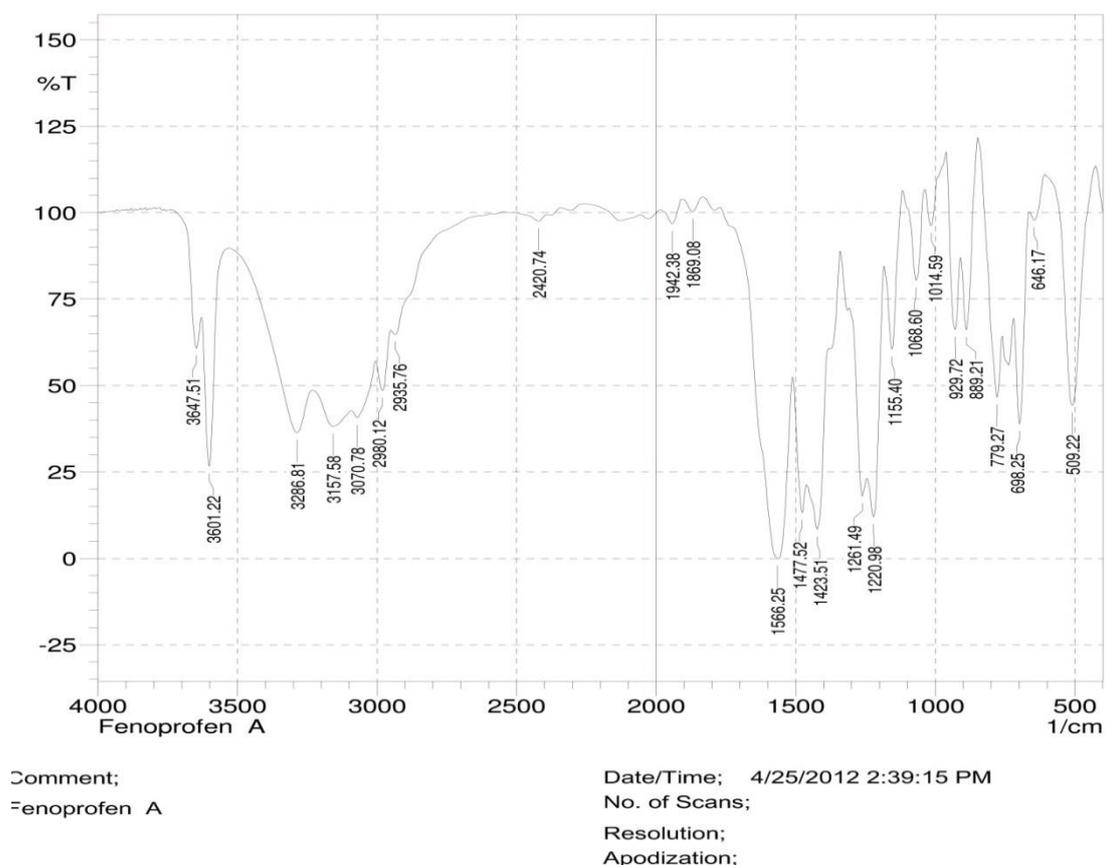
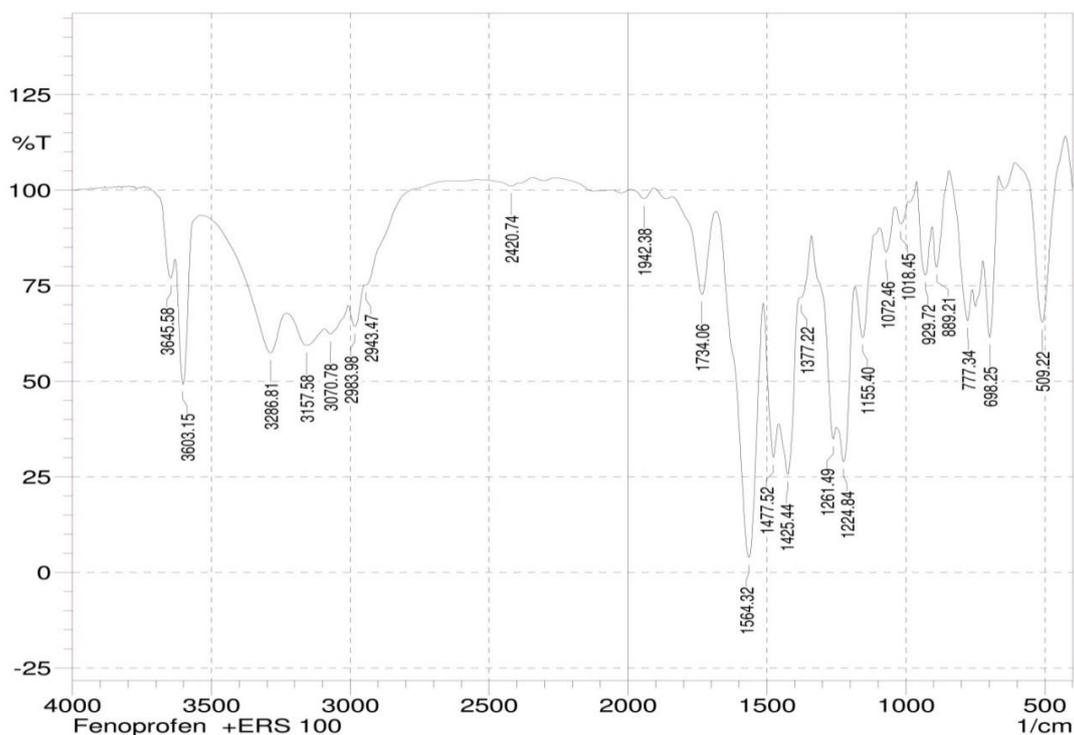


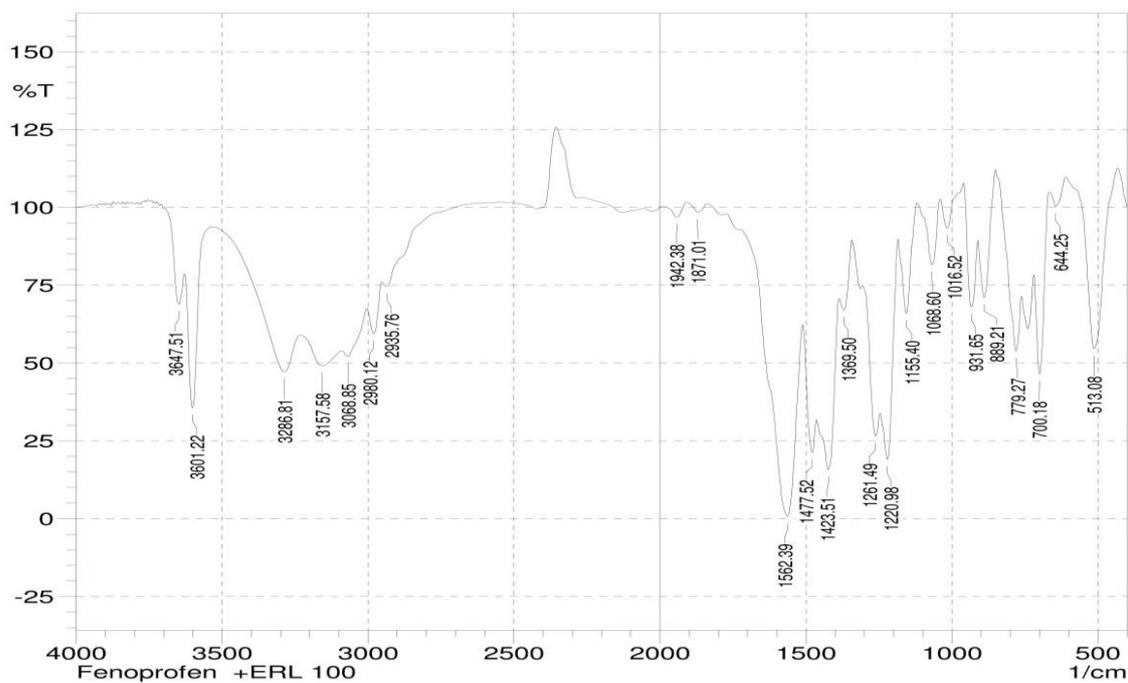
Figure 1: IR spectroscopy of fenopropfen alone.



Comment;
Fenopropfen +ERS 100

Date/Time; 4/25/2012 3:00:41 PM
No. of Scans;
Resolution;
Apodization;

Figure 2: IR spectroscopy of fenopropfen and ERS 100.



Comment;
Fenopropfen +ERL 100

Date/Time; 4/25/2012 3:06:24 PM
No. of Scans;
Resolution;
Apodization;

Figure 3: IR spectroscopy of fenopropfen and ERL 100.

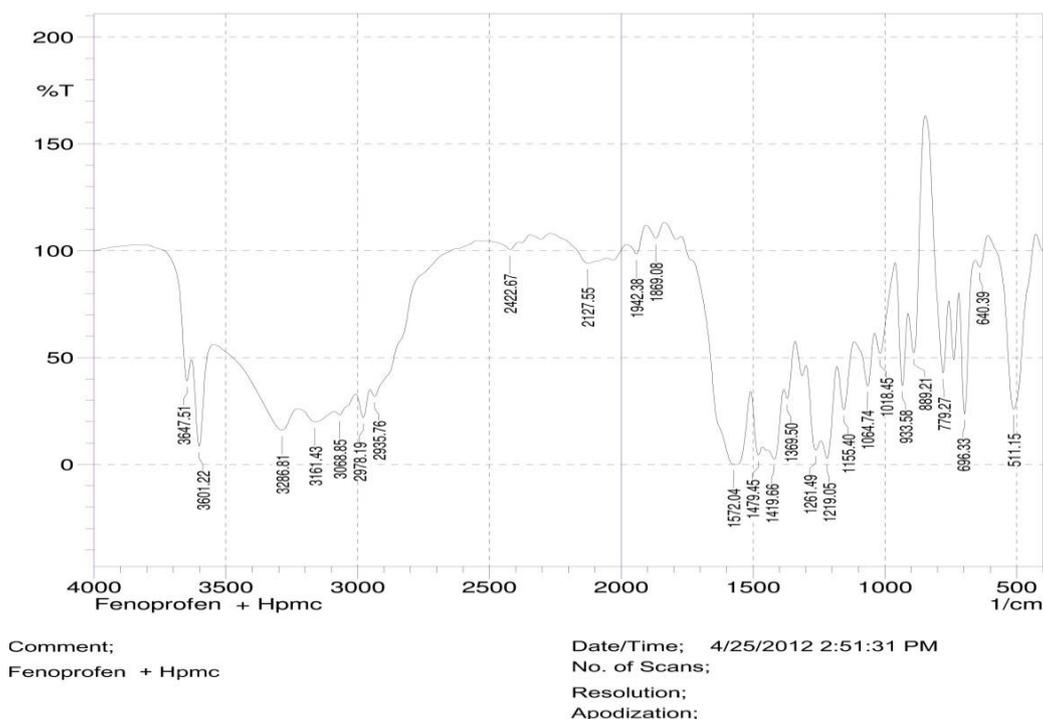


Figure 4: IR spectroscopy of fenopropfen and HPMC.

All the films were evaluated for their physical parameters and they were found to be smooth, flexible and transparent. They were also found to be uniform in their weight and thickness with low SD values. The drug content was found to range from 97.3% to 98.9% as shown in table 2.

Table 2: Physical parameters and drug content of the fenopropfen transdermal patches. HODS MATERIALS AND M

S.No.	Formulation Codes	Physical appearance	*Weight (mg)	*Thickness (mm)	**Drug content (%)
1	F1	++	275.3(0.45)	0.165(0.0067)	97.6
2	F2	++	274.8(0.35)	0.145(0.0044)	98.1
3	F3	++	278.3(0.38)	0.127(0.0022)	98.4
4	F4	++	290.8(0.44)	0.145(0.0044)	98.8
5	F5	++	293.5(0.35)	0.145(0.0044)	99.1
6	F6	++	273.6(0.25)	0.165(0.0067)	97.3
7	F7	++	274.8(0.34)	0.165(0.0067)	97.8
8	F8	++	277.7(0.45)	0.188(0.0044)	98.3
9	F9	++	270.8(0.32)	0.193(0.0067)	98.6
10	F10	++	272.5(0.48)	0.165(0.0067)	98.9

The figure inside the parenthesis denotes the standard deviation values

* → Average of five observations

** → Average of three observations

++ → Satisfactory.

The WVT studies for all monolithic formulations were conducted. The results are depicted in figure 5 and 6 for formulations from F1-F5 and F6-F5 respectively, which indicates that all formulations from F1 to F10 were permeable to water vapour. As shown in figure 5 and 6 it was observed that there was gradual increase in WVT for 7 days at regular intervals of 24 hrs that is the WVT rate constant was ranged between 3.307×10^{-2} to 3.352×10^{-2} gm/24hr/cm² for F1-F5 formulations and 3.05×10^{-2} gm/24hr/cm² for formulations F6-F10. The results revealed that the moisture permeability increased with increasing concentration of hydrophilic polymer (HPMC). The small moisture content in the formulations helps them to remain stable and from being completely dried and brittle film.

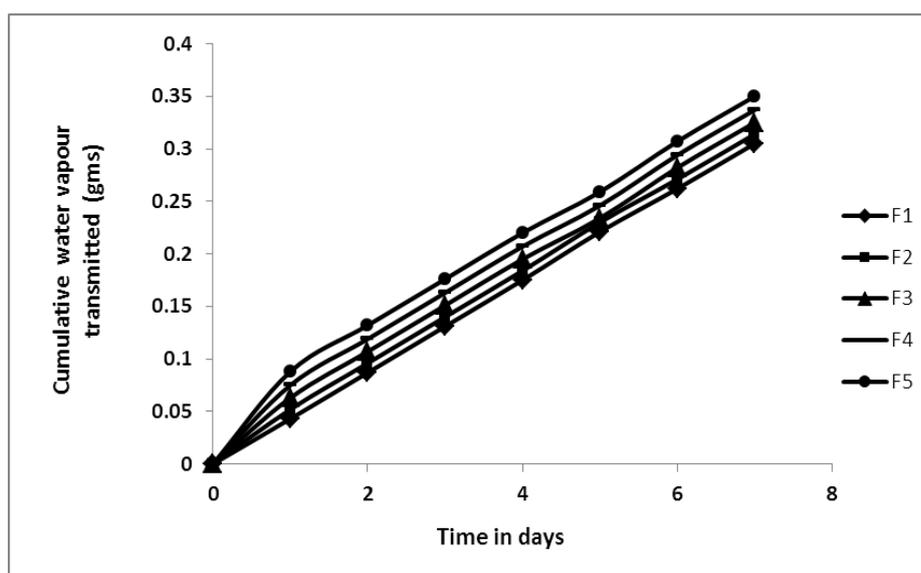


Figure 5: Water vapour absorption studies for monolithic systems F1 – F5.

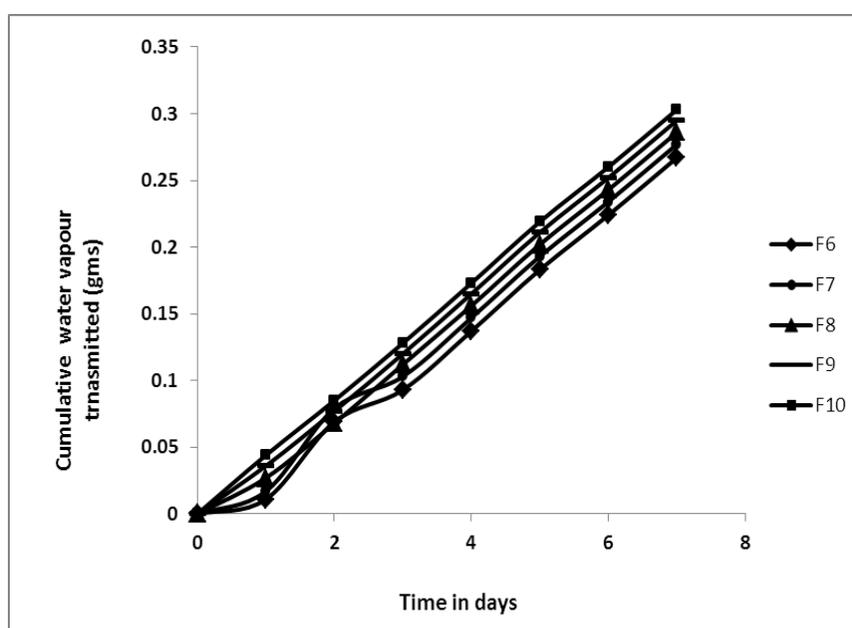


Figure 6: Water vapour absorption studies for monolithic systems F6 – F10.

Figures 7 and 8 shows the release profiles of fenoprofen from transdermal patches. Formulations F5 and F10 exhibited greatest percentage of drug release which were found to be 90.15% and 87.01%. The percentage of drug release from various ERL series F1, F2, F3, F4 were found to be 56.13%, 65.52%, 73.65% and 84.39% respectively, from ERS series F6, F7, F8 and F9 were found to be 51.13%, 59.64%, 71.35% and 82.62% respectively.

From the above results of *in vitro* drug release across rat abdominal skin, the addition of hydrophilic component (HPMC polymer) to an insoluble film former tends to enhance the drug release rates. As the proportion of HPMC increased in all the formulations, an increased drug release and permeation was observed.

Further the description of diffusion profiles by model function has been attempted using different kinetic model such as zero order, Higuchi square-root model and Korsmeyer-peppas model. The kinetic values and the slope values are depicted in table 3. The most appropriate model describing release with kinetic values ranging between 0.9676-0.9941 and n values 2.06 - 3.71 indicated that the drug release followed zero order kinetics and was found to be diffusion controlled, non-fickian release.

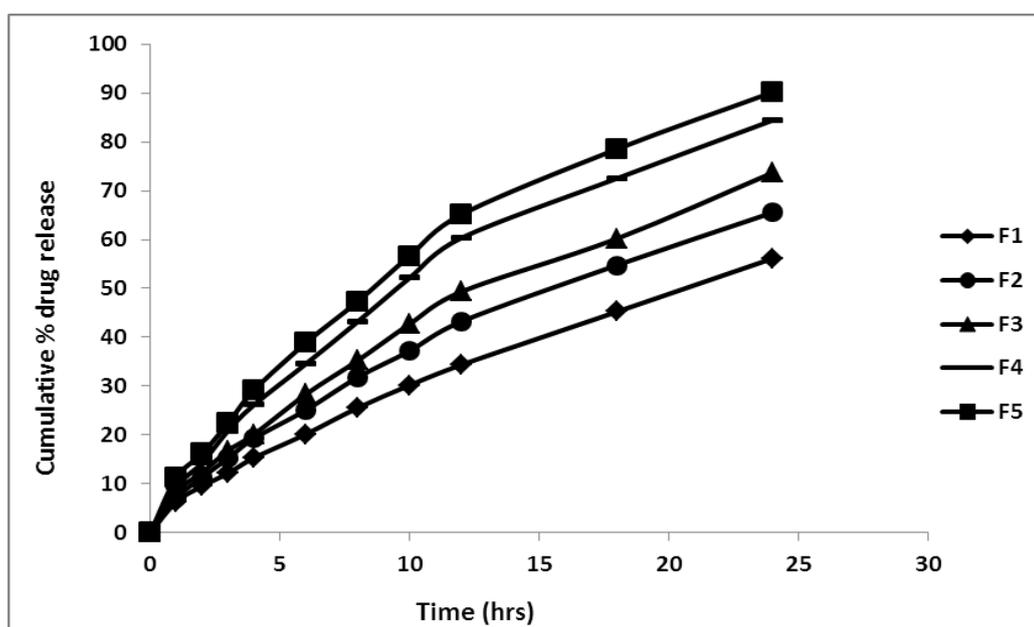


Figure 7: *In vitro* profiles of fenoprofen from monolithic systems of F1 – F5.

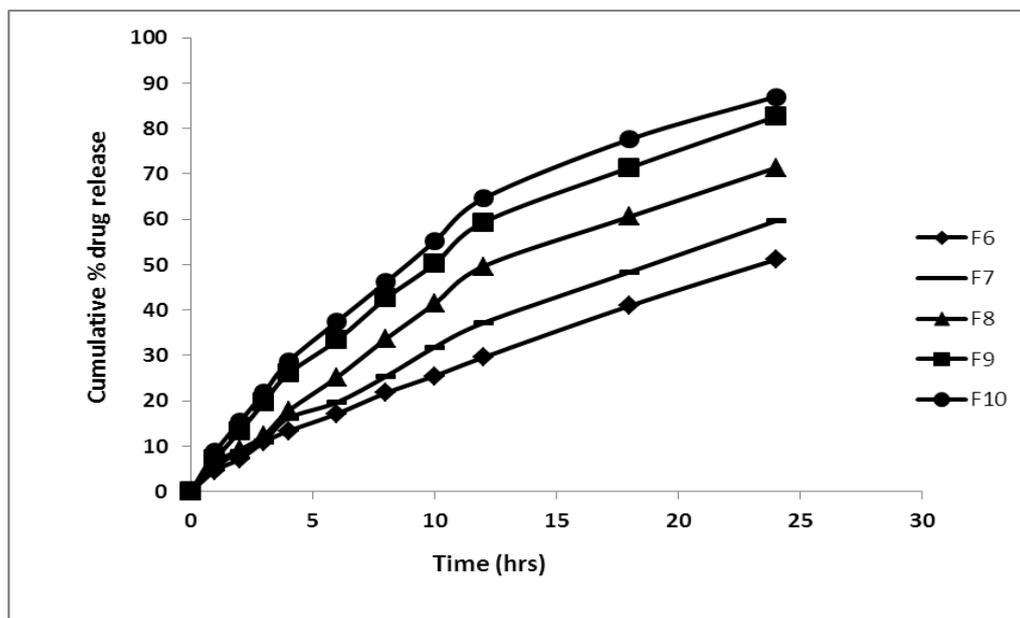


Figure 8: *In vitro* profiles of fenopufen from monolithic systems of F6 – F10.

The coefficient of determination indicated that the release data was best fitted with zero order kinetics and Higuchi equation (0.9676 – 0.9941) explained the diffusion controlled mechanism. The diffusion exponent n values of korsmeyer-peppas model was found to be in the range of 0.94 – 1.06 indicating non-fickian diffusion, super case II transport.^[14]

From the above *in vitro* drug release studies, formulations F5 and F10 showed good results in all aspects, hence these formulations were further subjected for *in vivo* studies.

Table 3: Comparative kinetic values of drug release from various transdermal patches.

S.No.	Formulation Codes	Zero order equation		Higuchi's equation		Korsmeyers-Peppas equation	
		n	r	n	r	n	r
1	F1	2.2531	0.9901	11.771	0.9863	0.9449	0.9106
2	F2	2.6691	0.9836	14.0907	0.9901	0.9802	0.9013
3	F3	2.9988	0.9827	15.8256	0.9888	1.0265	0.8943
4	F4	3.4936	0.9736	18.6475	0.9909	1.0265	0.8854
5	F5	3.7172	0.9707	19.9252	0.9921	1.0186	0.8702
6	F6	2.0689	0.9941	10.7042	0.9807	0.9663	0.9361
7	F7	2.4561	0.9912	12.7595	0.9818	1.0002	0.9295
8	F8	3.0635	0.9816	16.0538	0.9808	1.0642	0.9285
9	F9	3.4650	0.9731	18.4815	0.9896	1.0630	0.9034
10	F10	3.6579	0.9676	19.6405	0.9906	1.0494	0.8875

Finally, both the films (F5 and F10) were subjected for stability studies and observed for the changes in colour, appearance, flexibility and drug content at two different temperatures of

37⁰ C and 45⁰ C. The above observations were noted for one month at an interval of one week. All the systems were found to be stable at both the temperatures with respect to their physical parameters and drug content. The results are tabulated in table 4.

Table 4: Stability studies of F5 and F10 formulations at 37⁰ C.

S. No.	Time in days	Formulation Codes (Temperature 37 ⁰ C)		Formulation Codes (Temperature 45 ⁰ C)	
		F5	F10	F5	F10
		%drug remaining	%drug remaining	%drug remaining	%drug remaining
1	0	100	100	100	100
2	7	99.6	99.4	98.8	98.3
3	14	99.2	98.5	98.5	98.1
4	21	98.8	98.2	98.1	97.8
5	28	98.5	98.2	97.4	97.2

ACKNOWLEDGEMENT

The authors are thankful to VGST for providing grants to carry out the research work and also thankful to management and principal, Dayananda Sagar College of Pharmacy for their kind co-operation and support.

REFERENCES

1. Robert L Jackson, Marrow D Jason. Analgesic-antipyretic, anti-inflammatory agents and drugs used in the treatment of gout, 10th Edn., Mcgraw Hill publication., New York, 2003.
2. Chien Yie W. Concepts and system design for rate controlled drug delivery, 2nd Edn, Marcel Dekker Inc, 1992.
3. Chien YW, In; Robinson, J.R and Lee, V.H. Controlled drug Delivery: Fundamentals and Applications, 2nd Edn., Marcel Dekker Inc., Newyork, 1987; 523 - 524.
4. Alfonso RG, Remington: The Science and Practice of Pharmacy, 19th Edn, Mac publishing Company, Pennsylvania, 1985.
5. Pandey S, Praveen SH and Udupa N: Formulation and evaluation of Nimesulide Transdermal drug delivery systems. Indian J. Pharm. Sci, 62(5): 2000; 376-378.
6. Fregany A, Mohammed, Hussin Dhedr. Preparation and *in vitro/in vivo* evaluation of buccal adhesive properties of slow-release tablets containing miconazole nitrate. Drug Dev Ind Pharm, 2003; 29(3): 321-37.
7. Mundane BL., Evaluation of polymeric materials and screening of film coating agent. J Pharma Sci, 1964; 53: 395.

8. Raghavendra K, Doddayya H, Marihar SC, Patil CC, Habbu PV. Comparative evaluation of polymeric films for transdermal application. *The Eastern Pharmacist*, 2000; XLIII (153): 109-111.
9. Kusum devi V, Saisivam S, Maria GR, Deepti PU. Design and evaluation of matrix diffusion controlled transdermal patches of verapamil hydrochloride. *Drug Dev Ind Pharm*, 2003; 29(5): 495-503.
10. Gattani SG, Zawar LR, Kakade KN and Surana SJ. Optimization of transdermal films of lovastatin. *Indian Drugs*, 2008; 45(11): 883-889.
11. Flynn GL, Durrheim h, Higuchi WI. Permeation of hairless mouse skin II: Membrane sectioning techniques and influence on alkanol permeabilities. *J pharm Sci*, 1981; 70: 52-56.
12. Chien YW. Systemic drug delivery of pharmacologically active molecule across the skin. New York, 1991; 100: 100-102.
13. Narasimha MS, Mini Satish, Hamsa V. Drug release from terbutaline sulphate transdermal films across human cadaver skin. *Indian.J.Pharma. Sci*, 1997; 59(2): 75-76.
14. Peppas NA. Fitting controlled release and dissolution data. *Pharm. ActHelv*, 1985; 60: 110-111.