POMEGRANATE – A DOUBLE EDGED SWORD FOR THROMBOLYSIS AND OXIDATIVE STRESS

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ABSTRACT

The well recognized thrombolytic potential of the fruits of Punica granatum in vitro has influenced furthermore to confirm its clot lysing ability in vivo. In the current study, thrombolytic efficiency of the aril and rind of the fruits of pomegranate was analysed in the thrombus induced rat models. The ability of the extract to lyse the preformed clot was confirmed through the haematological profile, antioxidant status and histopathological examination of liver, heart and tail of experimental rats. The extracts of Punica granatum exhibited remarkable thrombolysis.

KEYWORDS: Thrombolysis; Punica granatum; Antioxidant; histopathology

INTRODUCTION

Acute myocardial infarction, stroke and venous thromboembolism that comprises cardiovascular diseases, can lead not only to sudden death but also to long-term disability at a large cost to society. Thrombus formed in the blood vessel can be dissolved with thrombolytic agents whose safety remains unanswered with several drawbacks such as systematic bleeding, short half-life and long term use is difficult.¹¹ The risks and hurdles associated with the currently used thrombolytic agents such as tPA
outweighs the purported benefits as thrombolytic agent, demanding an urge for sources with both antioxidant and thrombolytic property.

In most cases, it is not possible to obtain human samples from the site of thrombosis to study the disease and so it is advantageous for researchers to use animal models to investigate thrombosis in biological setting. The use of thrombosis models in rodents plays a crucial role in bridging the bench to bedside transition. The carrageenan induced tail thrombosis model in rats is useful for evaluating compounds having antithrombotic effects in drug discovery projects. The kappa carrageenan is a most potent thrombogen. As a consequence of thrombosis, tail infarction becomes visible few minutes after administration of carrageenan.

A surge in free radical production facilitating a vicious cycle that impinges on mitochondrial dysfunction, excitotoxicity, lipid peroxidation and inflammation is observed during ischemia as well as reperfusion. During reperfusion, oxidative stress reaches higher peaks and has a more sustained duration than other pathogenic mechanisms of ischemic cell death, further supporting the need for research of antioxidant therapies in reperfused individuals. Consequently, any plant source that has both clot lysing efficacy and excellent antioxidant potential will be the choice of interest since the fruit can overcome the ill effects caused by oxidative stress during reperfusion. Interestingly, pomegranate has well recorded antioxidant potential and its clot lysing efficiency is also proved in vitro. Therefore the present study was carried out with the aril and rind of pomegranate to ensure its thrombolytic efficiency in vivo.

MATERIALS AND METHODS
Collection and preparation of extracts
The fruit sample was collected from the Coimbatore city and certified by the Botanical Survey of India, Coimbatore. The voucher specimen was collected and maintained. The aril and rind of fresh fruits were collected, washed and homogenized using distilled water for the preparation of aqueous extract. It was then filtered using Whatmann No 1 filter paper and used for further study.

Evaluation of in vivo clot lysis in rat models:
In vivo studies were carried out by the oral administration of aril and rind of Punica granatum to examine the clot lysing ability and antioxidative effect to the experimental rats.
Maintenance of experimental animals

Male Swiss Albino rats were procured from Bangalore and were housed in microloan boxes in a controlled hygienic environment at temperature 25 ± 2°C and 12 hr dark/light cycle. The study was conducted after obtaining institutional animal ethical committee's clearance (IAEC No: KMCRET / Ph.D / 12/ 2014 – 15). As per the standard practice, the rats were segregated and quarantined for 15 days before the commencement of the experiment. They were fed on standard healthy diet and water ad libitum. The acute toxicity studies carried out confirmed that the extracts are not toxic even at the concentration of 5000 mg/kg.

Grouping of animals

The rats were divided into seven groups with six rats in each. The groupings are as follows:

<table>
<thead>
<tr>
<th>GROUPS</th>
<th>DESCRIPTION</th>
<th>EXTRACT / DRUG DOSE</th>
</tr>
</thead>
<tbody>
<tr>
<td>I</td>
<td>Control</td>
<td>-</td>
</tr>
<tr>
<td>II</td>
<td>κ – carrageenan only</td>
<td>-</td>
</tr>
<tr>
<td>III</td>
<td>κ – carrageenan + Streptokinase</td>
<td>30,000 IU</td>
</tr>
<tr>
<td>IV</td>
<td>κ – carrageenan + PgAA</td>
<td>250 mg/kg</td>
</tr>
<tr>
<td>V</td>
<td>κ – carrageenan + PgRA</td>
<td>250 mg/kg</td>
</tr>
</tbody>
</table>

PgAA – P. granatum Aril Aqueous; PgRA - P. granatum Rind Aqueous

Thrombus induction

Experimental rats were anaesthetized with i.p injection of ketamine hydrochloride (100 mg/kg). To attain effective clot formation κ – carrageenan (1 mg/kg) dissolved in saline was injected into the rat tail vein at a site 12 cm from the tip of the tail with a ligation. After a period of 10 minutes, the ligature was removed. The length of the infarct was monitored for thrombus formation. Once thrombus was formed, the animals were treated with respective extracts and monitored for the reduction in the length of the thrombus in rat tail for 30 days.

Haematological Parameters

At the end of the study, the rats were sacrificed after an overnight fasting. The blood of the animals was collected by heart puncture and the serum separated was used for the estimation of haematological parameters associated with thrombolysis.

<table>
<thead>
<tr>
<th>S.No</th>
<th>PARAMETERS</th>
<th>SOURCE</th>
<th>METHOD</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Haemoglobin</td>
<td>Citrated Blood</td>
<td>Drabkin and Austin, 1932\textsuperscript{[7]}</td>
</tr>
<tr>
<td>2</td>
<td>RBC</td>
<td></td>
<td>Sanderson and Phillips, 1981\textsuperscript{[8]}</td>
</tr>
<tr>
<td>3</td>
<td>Platelet</td>
<td></td>
<td>Automated Cell counter – CD1700</td>
</tr>
</tbody>
</table>
**In vivo antioxidant potential**

An oxidative stress is induced during clot formation and reperfusion, increasing the generation of free radicals. Thus, determining the tissue antioxidant status is essential to understand the effect of extracts on rats.

A part of the liver and kidney homogenate was prepared using PBS and used for the determination of enzymic and non-enzymic antioxidants. Antioxidants namely catalase, superoxide dismutase, glutathione peroxidase, vitamin C and reduced glutathione were then assessed in the liver and kidney of the experimental rats.

Catalase - Catalase activity in the tissue homogenate was assayed by the method of Sinha (1972).[9]

Superoxide dismutase - Superoxide dismutase activity in the tissue was determined by Kakkar (1984).[10]

Glutathione peroxidase - Glutathione peroxidase activity was determined by the method of Rotruck et al.[11]

Vitamin C - Ascorbic acid, a scavenger of oxyradicals was estimated by the method of Omaye et al., 1979.[12]

Reduced glutathione - Reduced glutathione, an intermediary redox metabolite in the ascorbate - glutathione cycle of scavenging hydrogen peroxide, was estimated according to the procedure described by Moron et al. (1979).[13]

**Histopathological examination**

Liver, heart and tail were removed from experimental animals and transferred to ice cooled containers with 10% formalin solution for histopathological observation (Cullin, 1979).[14]

**RESULTS AND DISCUSSION**

**Haematological parameters**

The pathology and physiology of man and animals when exposed to any foreign substance is first reflected in the blood components and the parameters usually analysed are haemoglobin, RBCs and WBCs. Blood form an important index for risk evaluation in both humans and rodents. So blood from all rats were analysed for components namely RBC, haemoglobin,
platelets, WBC, monocytes and neutrophils to study its association with thrombosis and thrombolysis. The results are tabulated in figure 1 and table 1.

**Figure 1**

**Level of RBCs, platelets and haemoglobin in experimental rats**

![Figure 1](image)

RBC is expressed as $10^6$ cells/µl,
Haemoglobin is expressed as g/dl,
Platelet is expressed as $10^5$ cells/µl

Total red blood cell count and haemoglobin levels revealed a similar pattern among all groups. It was found that the RBCs and haemoglobin increased when a thrombus is formed but the level remained well within the normal range. The primary thrombus formation is a critical phenomenon that involves activation of platelets and thereby platelet aggregation. RBCs that are generally regarded as innocent bystanders do play a significant role during thrombus formation and the mechanism of action needs further investigation.[15]

Platelet count was found to increase when a thrombus was formed and subsequently increased when treated with both the extract and standard streptokinase. The increase in the level of platelets when treated with the extracts were lower when compared with that of the drug, streptokinase. This marginal increase could be credited for the reoclusion, a major drawback in thrombolytic therapies. McRedmond et al.[16] reported that streptokinase activates platelets, thereby limiting its efficiency as a thrombolytic agent. In contrast, the level of platelets did not increase significantly when treated with fruit extracts, proving its safety and thrombolytic efficiency in vivo.
Table: 1 Level of WBCs, monocytes and neutrophils in experimental rats

<table>
<thead>
<tr>
<th>Groups</th>
<th>Treatments</th>
<th>WBCs x10^3 /μl</th>
<th>Neutrophils x10^3 /μl</th>
<th>Monocytes x10^3 /μl</th>
</tr>
</thead>
<tbody>
<tr>
<td>I</td>
<td>Control</td>
<td>11.65 ± 0.15</td>
<td>0.032 ± 0.002</td>
<td>2.95 ± 0.06</td>
</tr>
<tr>
<td>II</td>
<td>Thrombus induced</td>
<td>14.50 ± 0.21^a</td>
<td>0.05 ± 0.001</td>
<td>3.29 ± 0.07^a</td>
</tr>
<tr>
<td>III</td>
<td>Streptokinase Treated</td>
<td>13.50 ± 0.12^ab</td>
<td>0.044 ± 0.003</td>
<td>3.10 ± 0.11</td>
</tr>
<tr>
<td>IV</td>
<td>PgRA Treated</td>
<td>13.75 ± 0.16^ab</td>
<td>0.038 ± 0.001</td>
<td>2.85 ± 0.09^b</td>
</tr>
<tr>
<td>V</td>
<td>PgAA Treated</td>
<td>13.45 ± 0.17^ab</td>
<td>0.039 ± 0.004</td>
<td>2.91 ± 0.05^b</td>
</tr>
<tr>
<td>Normal values</td>
<td>6.6 – 12.6</td>
<td>0.01 - 0.04</td>
<td>1.77 – 3.38</td>
<td></td>
</tr>
</tbody>
</table>

PgRA – Punica granatum rind aqueous
PgAA – Punica granatum aril aqueous

An increase in total WBC count and neutrophils was found in thrombus induced groups while they fell towards normalcy when treated with streptokinase and fruit extracts. The level of monocytes was within the normal range among the groups that were analysed.

Literatures support the fact that there is WBC infiltration in the first 12 hours after myocardial infarction, thereby promoting tissue damage. Following ischaemia, neutrophils are involved in the inflammatory response. Barbui et al.[17] have reported that leukocytosis; an increase in WBC count appears to be a potential threat factor for thrombosis. An elevation of WBC count during acute myocardial infarction is associated with adverse outcomes such as reduced epicardial blood flow, thromboresistance, new congestive heart failure and even death. A systemic inflammatory reaction is induced due to thrombus formation and consequent vein wall damage in deep vein thrombosis patient and as a consequence, increased level of WBCs was observed.[18]

Antioxidant potential

A surge in free radical production is observed during ischemia/reperfusion resulting in a further increase in oxidative stress and tissue damage. During reperfusion, oxidative stress reaches higher peaks and has a more sustained duration than other pathogenic mechanisms of ischemic cell death, further supporting the need for research of antioxidant therapies in reperfused individuals.[6]

Therefore, the risks and hurdles associated with the currently used thrombolytic agents such as tPA overshadows the supposed benefits as thrombolytic agent, demanding an urge for sources with both antioxidant and thrombolytic property. Therefore, the effect of Punica
granatum in the antioxidant status of animals was determined in kidney and liver. Activity of enzymic antioxidants namely catalase, superoxide dismutase and glutathione peroxidase and levels of non-enzymic antioxidants such as vitamin C, vitamin E and reduced glutathione were evaluated and the results are as tabulated in tables 2 and 3.

From the results it was observed that the fruit extract conferred good antioxidant protection against the oxidative stress that was found to be peaked during the thrombus formation and lysis. A significant increase in the levels of non-enzymic antioxidants and activity of enzymic antioxidants were noted in animals treated with Punica granatum pulp extracts than rind extracts.

Nekooeian[19], showed that the pomegranate seed oil decreased the oxidative stress without affecting the serum lipid profile. Moneim et al.[20] reported a significant increase in superoxide dismutase and catalase activities of liver and kidney samples of rats that received pomegranate. Chalfoun-Mounayar et al.[21] reported that pomegranate molasses possesses a powerful antioxidant activity with increased superoxide dismutase activity.
Table: 2 Activity of enzymic antioxidants in experimental rats

<table>
<thead>
<tr>
<th>Groups</th>
<th>CAT (U/g protein)</th>
<th>SOD (U/g protein)</th>
<th>GPx (U/g protein)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Kidney</td>
<td>Liver</td>
<td>Kidney</td>
</tr>
<tr>
<td>I</td>
<td>813.55 ± 25.33</td>
<td>743.63 ± 34.83</td>
<td>346.44 ± 23.26</td>
</tr>
<tr>
<td>II</td>
<td>632.23 ± 25.76</td>
<td>541.65 ± 24.81</td>
<td>267.34 ± 25.76</td>
</tr>
<tr>
<td>III</td>
<td>710.21 ± 35.47</td>
<td>579.76 ± 37.19</td>
<td>375.59 ± 41.03</td>
</tr>
<tr>
<td>IV</td>
<td>889.95 ± 20.45</td>
<td>882.90 ± 12.24</td>
<td>417.16 ± 14.66</td>
</tr>
<tr>
<td>V</td>
<td>923.20 ± 20.28</td>
<td>964.00 ± 26.17</td>
<td>372.38 ± 19.52</td>
</tr>
</tbody>
</table>

CAT – Catalase  
SOD – Superoxide Dismutase  
GPx – Glutathione Peroxidase

Table: 3 Level of non-enzymic antioxidants in experimental rats

<table>
<thead>
<tr>
<th>Groups</th>
<th>VITAMIN E (μg/ g tissue)</th>
<th>VITAMIN C (mg/ g tissue)</th>
<th>REDUCED GLUTATHIONE (nmoles/g tissue)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Kidney</td>
<td>Liver</td>
<td>Kidney</td>
</tr>
<tr>
<td>I</td>
<td>2.78 ± 0.07</td>
<td>2.41 ± 0.05</td>
<td>0.22 ± 0.001</td>
</tr>
<tr>
<td>II</td>
<td>1.85 ± 0.06 (^a)</td>
<td>2.04 ± 0.03 (^a)</td>
<td>0.18 ± 0.005</td>
</tr>
<tr>
<td>III</td>
<td>1.75 ± 0.09 (^a)</td>
<td>2.12 ± 0.04 (^a)</td>
<td>0.15 ± 0.006</td>
</tr>
<tr>
<td>IV</td>
<td>3.14 ± 0.11 (^abc)</td>
<td>3.08 ± 0.09 (^abc)</td>
<td>0.25 ± 0.009</td>
</tr>
<tr>
<td>V</td>
<td>3.52 ± 0.13 (^abc)</td>
<td>3.49 ± 0.12 (^abc)</td>
<td>0.29 ± 0.013</td>
</tr>
</tbody>
</table>
A sensible strategy and a herculean task that must be adopted before a costly translational research is to explore newer antioxidant drug which must show recognisable additive or synergistic effects with thrombolytic agents in thromboembolic models.\textsuperscript{[22]} The results of the present study confirm the same and collectively signify that the fruit Punica granatum has an augmentive effect on thrombolysis by rendering good oxidative protection with its numerous antioxidants.

**HISTOPATHOLOGICAL CHANGES IN EXPERIMENTAL RATS**

Histopathological examinations of liver, heart and tail of control and experimental rats of the various groups studied was carried out to test the cytotoxicity and clot lysing effect of the aril and rind of Punica granatum. The cellular changes are indicated in figures 2, 3, 4 and the findings are discussed below.

Altogether, there was no major change in histopathology of the tissues in the treated groups confirming that the extracts did not render any cytotoxic effect. Moreover, the tail histology validated the thrombolytic efficiency of the fruit Punica granatum.

Alferah in 2012\textsuperscript{[23]} studied the toxicity induced by Lawsonia inermis leaf extract and the subsequent histological changes in liver and kidney of male wistar rats. Moderate degenerative changes in epithelium lining in renal tubes and distortion and degeneration of hepatocytes were observed. No pathological observations were made when a medicinal plant extract (Ankaferd Blood Stopper) was applied in deep tissue injury in rats.\textsuperscript{[24]}

![Group I](image)
Group I – Histology of liver sections of control animals showed normal lobular architecture. The hepatocytes were intact with no periportal inflammation and the portal triad was normal.

Group II – The hepatocytes maintained normal lobular architecture with mild portal tract inflammation. Microvesicular steatotic hepatocytes were also seen with no cholestralosis.

Group III – The sections showed pale steatotic area with central vein and extensive microvesicular fatty change.

Group IV – The cells showed mild alterations in the lobular architecture with very mild inflammation.

Group V – There were no alterations in the architecture in the liver section. Only a mild inflammation was detected.
Group I – Histological sections of heart showed normal myocardium with no inflammation or necrosis.

Group II – The myocardium showed congested veins and mild chronic inflammation. Anisonucleosis and nucleomegaly of myocardium were also observed.

Group III – Congested vessels in myocardium and few necrotic muscle fibres were observed.

Group IV – The cells showed congested vessels with normal myocardium.

Group V – There were no alterations in the architecture in the heart section. The myocardium was normal with no chronic inflammation.
Figure: 4 Histopathological observations of tail of experimental animals

Group I – Histological sections of tail showed normal epidermis, dermis with normal blood vessels and hair follicles.
Group II – The histopathological observations of tail showed congested blood vessels with edema and mild inflammation. Thick and thin walled vessels with fibrocollagenous stroma were observed.
Group III – Normal blood vessels, dermis and epidermis were observed. The dermis showed normal proliferation of blood vessels.
Group IV – The skin showed congestion with edema and congested blood vessel.
Group V – There was no evidence of toxic changes. There was slight lymphocyte infiltration.

Literatures reveal that urokinase plasminogen activator; a thrombolytic drug activates oxidative stress. Supplementation with pomegranate juice reduces the effect of thrombosis and it shows plasminogen activation. With a plethora of shortcomings and negativity of the available thrombolytic therapies, these modest findings of the present study gain significance in employing the fruit Punica granatum for efficient clot lysis.

CONCLUSION

Oxidative stress is the key root for several lifestyle associated diseases in today’s world. Pomegranate, which was extensively exploited for its antioxidant potential has now been recognized as a double edged sword which has potent thrombolytic efficiency as well.
However, further investigations are warranted to have a profound insight into the mechanism of action of the fruit for thrombolysis.

REFERENCES


